

1 **Development of aqueous protein/polysaccharide mixture-based inks for 3D**
2 **printing towards food applications**

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26 **ABSTRACT**

27 Three-dimensional (3D) food printing is a promising technique as it allows to create elaborated
28 food constructs or customized food for elderly people and other sensitive population (i.e. children,
29 pregnant women or athletes). The structural and textual properties of food constructs depend
30 strongly on the ingredients used and the shape produced. Two important biological
31 macromolecular components, namely protein and polysaccharides, are the most important in terms
32 of nutritional content. In this work, we developed a water-based protein/polysaccharide food ink
33 from a mixture of gelatin B (GB) and xanthan gum (XG) for 3D printing. A 40-layer 3D scaffold
34 with defined features was successfully printed at room temperature using an aqueous mixture of 3
35 wt% GB and 10 wt% XG. Adding 37.5 mM calcium ions (Ca^{2+}) or lowering the storage
36 temperature to 4 °C improved the rheological properties of the ink, which endowed the scaffold
37 with higher shape fidelity right after printing and good shape retention for at least 96 h. Texture
38 profile results revealed that the infill percentage of the scaffold is critical to textural properties of
39 the scaffold, highlighting the advantage of 3D printing in tuning sensory food perception.

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42 **Keywords:** 3D food printing; Gelatin; Xanthan gum; calcium salt; rheological properties; textural
43 profile results

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53 **1. Introduction**

54 Three-dimensional (3D) food printing is an emerging manufacturing technology using the additive
55 principle through computer-controlled layer-by-layer deposition of materials (Liu, Zhang,
56 Bhandari, & Yang, 2018). It enables the production of customized food designs and creative food
57 flavors with personalized nutrition according to individual health conditions (Lille, Nurmela,
58 Nordlund, Metsä-Kortelainen, & Sozer, 2018; Sun, Zhou, Huang, Fuh, & Hong, 2015).
59 Additionally, 3D food printing technology is a convenient production process (H. W. Kim, Bae,
60 & Park, 2017). It allows to fabricate complex food design with precise temperature controlled
61 during the printing process (Sun, Zhou, Yan, Huang, & Lin, 2018) and with relatively less material
62 loss (Vancauwenberghe, et al., 2017). Therefore, it has the potential to replace some of the more
63 cumbersome processes of traditional food production methods (Chen, et al., 2019; H. W. Kim, et
64 al., 2017).

65 The first reported 3D printer goes back to 2007 at Cornell University where the researchers
66 developed the “Fab@Home” model (Gross, Erkal, Lockwood, Chen, & Spence, 2014). Since then,
67 researchers have developed different printing techniques, such as extrusion-based printing,
68 selective laser sintering and inkjet printing etc.(Wegrzyn, Golding, & Archer, 2012). Direct-ink
69 writing (DIW) is a 3D printing technique employing a computer-controlled translation stage,
70 which moves an ink-deposition micro-nozzle, to generate materials with controlled design and
71 composition. It is an adaptable approach which offers flexibility requirement for soft materials,
72 such as colloidal suspensions, fugitive ink and hydrogels. So far, several food inks including
73 chocolate (Lanaro, et al., 2017), pasta (Godoi, Prakash, & Bhandari, 2016), mushroom (Keerthana,
74 Anukiruthika, Moses, & Anandharamakrishnan, 2020), cheese (Le Tohic, et al., 2018) and fish gel
75 (Lin Wang, Zhang, Bhandari, & Yang, 2018) have been successfully printed using the DIW
76 technique. Different mechanisms have been employed to facilitate shape retention during post
77 extrusion, including solvent evaporation, gelation, temperature control and UV polymerization.
78 However, there is still a lack of fundamental knowledge on 3D printing of essential constituents
79 of food (H. W. Kim, et al., 2017; Liu, et al., 2018; Sun, Peng, et al., 2015).

80 Proteins and polysaccharides are two important biological macromolecules in food systems, which
81 perform complementary nutritional, structural and textural functions (C. S. Wang, Virgilio, Wood-
82 Adams, & Heuzey, 2017; X.-Y. Wang, Wang, & Heuzey, 2015). Attractive (biopolymers with

83 opposite charges) and repulsive interactions (biopolymers with same or no charge) can occur
84 depending on the environmental pH and ionic strength (Zeeb, Thongkaew, & Weiss, 2014).
85 Electrostatic forces and hydrogen bonding are usually the dominant interactions between the two
86 components, while other weak interactions should also be considered such as van der Waals forces
87 and hydrophobic interactions (McClements, 2006). Different functional attributes can be provided
88 by these interactions, such as the ability to thicken solutions, form gels or emulsions (Tolstoguzov,
89 1995).

90 Gelatin, a linear protein, is obtained by either acid (type A) or alkaline (type B) hydrolysis of the
91 parent protein collagen, which is derived from animal skin and/or bones (Williams & Phillips,
92 2003). It has a helix-to-coil transition temperature, below which the random coils assemble into
93 triple helices, resulting in gelation via forming a 3D network (Joly-Duhamel, Hellio, Ajdari, &
94 Djabourov, 2002). Xanthan gum (XG) is an anionic food-grade polysaccharide produced by
95 aerobic fermentation, which is composed of a β -(1-4)-D-glucose main chain and a trisaccharide
96 branch (D-glucose, D-mannose and D-gluconate) as side chains (Pelletier, Viebke, Meadows, &
97 Williams, 2001). The XG molecules undergo a thermal-reversible coil-helix transition in solution,
98 and the transition temperature can be shifted to higher temperatures by the addition of electrolytes
99 (Song, Kim, & Chang, 2006) or proteins (C.-S. Wang, Virgilio, Wood-Adams, & Heuzey, 2018).
100 XG is widely used in food engineering owing to its shear-thinning properties as well as the
101 resistance against heat, acids, bases, salts and even enzymes (C.-S. Wang, Natale, Virgilio, &
102 Heuzey, 2016; C.-S. Wang, Virgilio, Carreau, & Heuzey, 2021). Moreover, required food
103 properties can be easily adjusted by XG addition, such as viscosity, texture appearance, etc.
104 (Kumar & Saini, 2021; Salehi, 2020). Although XG is a non-gelling polysaccharide, it can form a
105 gel in the presence of trivalent ions or when mixed with other polysaccharides, or even proteins
106 (Bertrand & Turgeon, 2007; Le & Turgeon, 2013; C.-S. Wang, et al., 2021; C.-S. Wang, et al.,
107 2018; C. S. Wang, et al., 2017).

108 Several food inks have been developed so far. For example, gel formed by low methoxylated pectin
109 (an anionic polysaccharide) has been used to print edible objects with DIW (Vancauwenberghe, et
110 al., 2017). The printed 3D structure can be greatly stabilized with the addition of CaCl_2 during and
111 after printing, due to ion bridging effects. The 3D printability of a mixture of gelatin and kappa-
112 carrageenan was investigated by Warner et al. (Warner, Norton, & Mills, 2019). It was found that

113 the addition of kappa-carrageenan enabled better control of the printing process compared to pure
114 gelatin and they attribute it to the increase of the gelling temperature.

115 Synergistic gelation properties have been previously reported for gelatin B (GB) with XG in
116 aqueous solution at a pH around the isoelectric point of GB (pI~5.3) (C.-S. Wang, et al., 2016). In
117 addition, the gel preparation process does not involve thermal, enzyme or any other denaturing
118 treatment, which is favorable for protecting micronutrients in food. It provides a promising way to
119 fabricate group-specific food. However, up to now, there is no specific study in the literature on
120 3D printing of GB/XG mixed gels with careful parameter control and detailed structure analysis.

121 In this work, we aim to shed light on the 3D printing of a protein/polysaccharide mixed gel system
122 for food application. 3D printing of a mixture of GB and XG based on the DIW method is presented
123 and analyzed in terms of the interactions between GB and XG. The effects of Ca^{2+} addition and
124 storage temperature after printing on shape retention, gel strength and textural properties are
125 investigated in order to clarify the printed object quality.

126

127 **2. Materials and methods**

128 **2.1. Materials**

129 Gelatin (type B, G6650, bloom index ~225, molecular weight: 20-25 kDa, pI = 4.7-5.3) and CaCl_2
130 $\cdot 2\text{H}_2\text{O}$ were purchased from Sigma-Aldrich Canada. XG (KELTROL RD, food-grade quality,
131 molecular weight: $\sim 10^3$ kDa) was supplied by CP Kelco U.S. All other reagents and chemicals
132 were of analytical grade and used without further purification.

133 **2.2. Food ink preparation**

134 GB solutions (3 wt%) were prepared by allowing the GB to swell in Milli-Q water (18.2 Ω) for
135 15-20 min, followed by gentle magnetic stirring at 40 °C for 15 min. The solutions were used on
136 the same day. 10 wt% XG dry powder was then added to the GB solution. The GB/XG mixed
137 solution was mechanically stirred at a speed of 600-700 rpm for at least 12 h at 40 °C to ensure
138 complete dissolution. The final pH of prepared solution was 5.8. For comparison purposes, a neat
139 XG solution of 10 wt% concentration was also prepared. Ca^{2+} was added to the GB/XG aqueous
140 mixture at concentrations that varied from 0, 12.5, 37.5, 62.5 to 75 mM, within the recommended

141 intake by the Institute of Medicine (IOM) (Ross, et al., 2011). The pH did not change with Ca^{2+}
142 addition. The ink was then centrifuged (5000 rpm, 1 h) at room temperature (22 °C) to eliminate
143 air bubbles.

144 ***2.3.3D printing process and 3D scaffold fabrication***

145 The food ink was inserted into a syringe (3 mL, Nordson EFD) linked with a cylindrical
146 micronozzle (exit inner diameter $D = 200 \mu\text{m}$, capillary length $L = 12.7 \text{ mm}$, Nordson EFD). Then
147 the loaded syringes were fixed to a high-pressure adaptor (HP-7x Nordson EFD), which was
148 installed on a computer assisted 3-axis positioning stage (I&J2200-4, I&J Fisnar Inc.). The 40-
149 layer 3D scaffold was printed by depositing filaments in a layer-by-layer manner with the GB/XG
150 food ink, as well as with different concentrations of Ca^{2+} solution. All scaffolds were printed at a
151 speed of 7 mm/s and under appropriate applied pressures at room temperature ($22 \pm 1 \text{ }^\circ\text{C}$). The
152 speed of 7 mm/s was chosen based on preliminary experiments as a compromise between shape
153 fidelity and printing time. Specific applied pressures are reported in the legend of **Figure S1**.

154 ***2.4.Rheological measurements***

155 In the low shear rate region ($0.01\text{-}10 \text{ s}^{-1}$), steady-shear measurements of GB/XG aqueous mixtures
156 were performed using a rough surface parallel plate flow geometry with 25 mm diameter with a
157 stress-controlled rheometer (MCR-502, Anton Paar, Graz, Austria). The gap between the plates
158 was 1 mm. A rough surface was chosen to prevent wall slip. Mineral oil was used to cover the
159 edge of the sample in the parallel plate flow geometry to prevent water evaporation. The
160 measurements were carried out at room temperature (22 °C).

161 In the high shear rate region, the process-related viscosity was calculated by capillary flow analysis
162 (CFA) (Bruneaux, Therriault, & Heuzey, 2008; Guo, Heuzey, & Therriault, 2014). Different food
163 inks were loaded into the syringe (3 mL, Nordson EFD). A 60 mm filament was extruded onto a
164 glass substrate through a micronozzle (exit inner diameter $D = 200 \mu\text{m}$, capillary length $L = 12.7$
165 mm, Nordson EFD) at a speed of 0.5 mm/s for 120 s with each food ink. The different pressures
166 were set under a range between 70 and 900 kPa. The weight of each extruded filaments was then
167 measured with a high-precision balance (GH-202, A&D Engineering) at room temperature, which
168 was used to calculate the process-related viscosity according to the method reported by Bruneaux
169 et al.(Bruneaux, et al., 2008).

170 The effects of Ca^{2+} and storage temperature after printing on the dynamic viscoelastic properties
171 of the food inks were also investigated using the same geometry. Linear viscoelastic (LVE) regime
172 was determined by strain sweeps in the range from 0.01 % to 100 % at angular frequency = 1 rad/s.
173 Frequency sweeps were performed over a range of 0.1 to 10 rad/s within the identified LVE regime
174 (strain = 0.2-5%). The storage modulus (G') and loss modulus (G'') were recorded using
175 Rheocompass software (Anton Paar).

176 For samples stored at 4 °C, the lower plate was pre-cooled down at 4 °C using the temperature-
177 controlled Peltier system before the test. All the rheological experiments were performed three
178 times.

179 ***2.5. Morphological characterization***

180 Optical images of 2D and 3D structures were captured using an optical microscope (BX-61,
181 Olympus) and a digital camera (EOS TRBRL T4I, Canon). The images were then analyzed with
182 Image Pro Plus 6.0 (Media Cybernetics).

183 ***2.6. Determination of dimensional stability of 3D-printed structures***

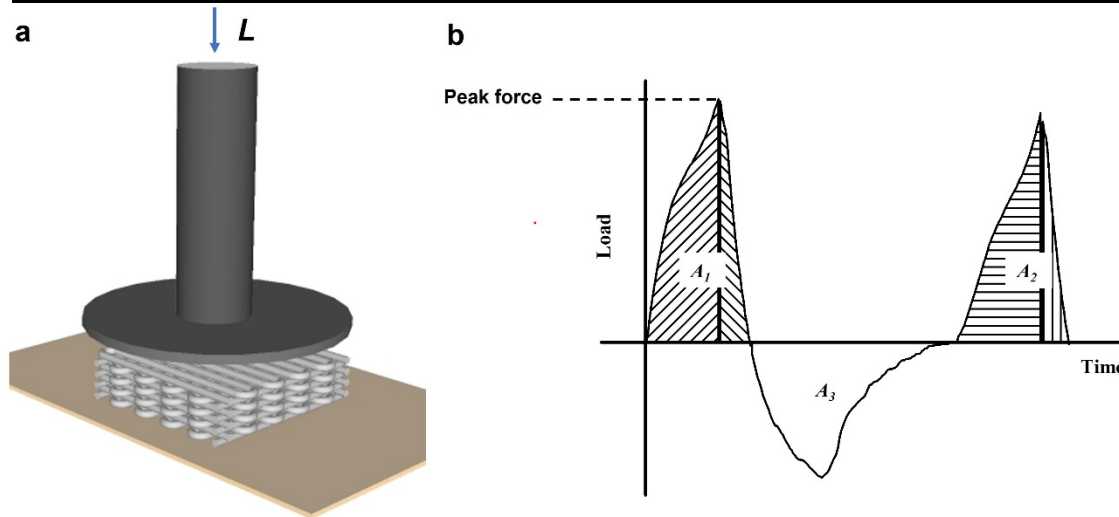
184 The printed dimensional stability obtained using the different food ink formulations was evaluated
185 by directly following the thickness variation of the as-printed 3D scaffolds up to 96 h. The images
186 of the samples were recorded 0, 24 and 96 h after printing, respectively. The thickness of the
187 samples was measured using a caliper. All samples were put in petri dishes and then stored under
188 a hood in order to maintain constant room temperature and humidity, or in a refrigerator at 4 °C.

189 ***2.7. Compression tests for textural profile analysis***

190 Mach-1TM Micromechanical tester (Biomomentum Inc.) was used for a double-cycle compression
191 tests to obtain load-time curves. A 25 mm diameter cylindrical probe was used to compress the
192 printed 40-layers 3D scaffolds (18 mm × 18 mm) composed of GB/XG food inks with different
193 Ca^{2+} contents and different infill levels (**Figure 1a**). The main test setting parameters are listed in
194 **Table 1**. Triplicate measurements of samples were carried out. **Figure 1b** shows how the various
195 properties of the textural profile test are measured and calculated. More explanations on the various
196 properties measured are given in the Results and discussion section.

197 **Table 1.** Parameters used for the compression tests

Setting parameters	Value	Unit
Contact force	1	g
Contact speed	0.1	mm/s
Test speed	0.5	mm/s
Withdrawal speed	0.5	mm/s
Compression distance	2	mm



198

199 **Figure 1.** (a) Schematic representation of the compression test using the Mach-1™
200 Micromechanical tester (b) Textural profile diagram for gels. (Hardness: Peak Force;
201 Cohesiveness: A_2/A_1 ; Adhesiveness: A_3 ; Gumminess: Hardness \times Cohesiveness.)

202 2.8. Statistical analysis

203 Triplicate tests have been performed for all the tests mentioned above. The mean of the results \pm
204 standard deviation (SD) was used to present the data and analyzed by SPSS software (SPSS 20.0,
205 IBM, Chicago, IL, USA). Significant differences ($p < 0.05$) between mean values of samples were
206 determined using Duncan's Multiple Range test.

207

208 3. Results and discussion

209 3.1. 3D printed structures

210 **Figure 2a** and **2b** show the 3D printing process using the food ink based on GB/XG aqueous
211 mixture and the scaffold model used, respectively. The effect of salt addition is observable when
212 XG concentration is higher than 0.3% (Chang, Im, Prasadhi, & Cho, 2015; de Moura & Moreno,

213 2019; Zatz & Knapp, 1984). Considering a gel which can flow under applied pressure and maintain
214 its structures after deposition, 10 wt% XG has been selected. Hence, a formulation of ink with 3
215 wt% GB and 10 wt% XG aqueous mixture has been obtained after carefully adjusting the
216 composition. It allowed printing a scaffold with accurate features (**Figure 2c**). In comparison, the
217 scaffold printed with a neat 10 wt% XG aqueous solution does not retain its structure after
218 deposition, exhibiting a poor self-supporting behavior (**Figure S1a**). Hence, it is clear that the
219 addition of GB imparts improved printability to the food ink. The addition of Ca^{2+} enhances the
220 shape retention of the scaffold printed by the food ink of GB/XG mixtures, as shown by the sharper
221 edges (**Figure 2c and 2d**).

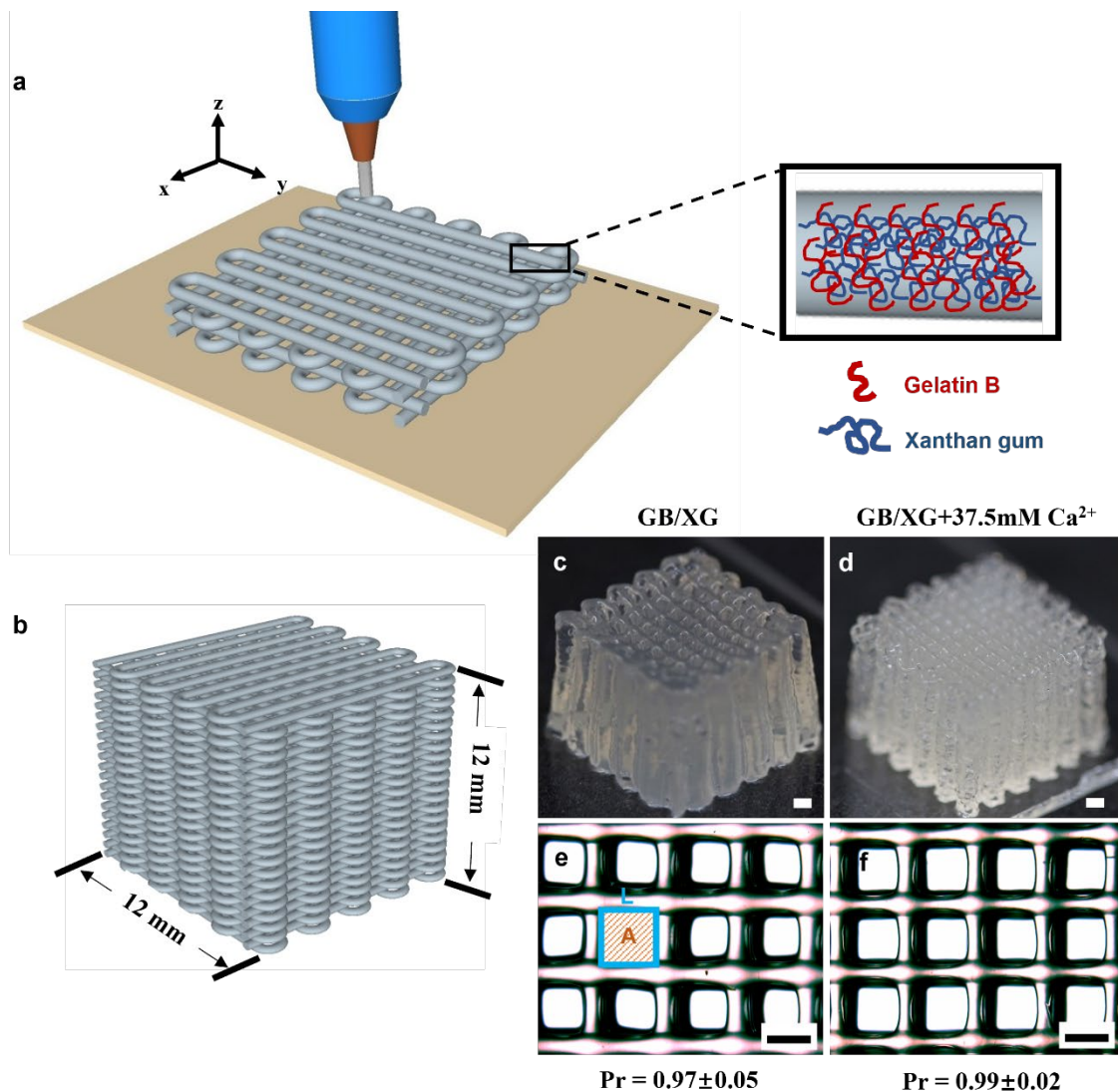
222 The top-view printability (Pr) (Equation 1) defined by Ouyang et al. (Ouyang, Yao, Zhao, & Sun,
223 2016) is used to quantitatively describe and compare the gelation condition and shape fidelity of
224 the food inks. The Pr is calculated based on the circularity (C) (Equation 2) estimated by the
225 perimeter L and pore area A (schematically shown in **Figure 2e**). When the Pr value is equal to 1,
226 the interconnected channels of the filaments show a square shape, indicating an ideal gelation.
227 Larger Pr value presents a greater gelation degree. Otherwise ($Pr < 1$), a lower gelation degree is
228 noted. **Figure 2e and 2f** show the top-view images of the as-printed GB/XG scaffold with 0 and
229 37.5 mM of Ca^{2+} along with their Pr values. These values show that the printability of both food
230 inks is within the proper range, while the one with Ca^{2+} addition is slightly closer to 1, indicative
231 of better printability with this content of Ca^{2+} in the GB/XG food ink.

$$232 \quad Pr = \frac{\pi}{4} \cdot \frac{1}{C} = \frac{L^2}{16A} \quad (1)$$

$$233 \quad C = \frac{4\pi A}{L^2} \quad (2)$$

234 However, 62.5 mM Ca^{2+} addition (**Figure S1d**) causes irregular extrusion with structure defects.
235 The poor structure quality is probably due to an over-gelation of the printed filament. Among the
236 tested Ca^{2+} concentrations, the scaffold with 37.5 mM Ca^{2+} shows the best appearance (**Figure 2d**
237 **and Figure S1c**).

238



239

240 **Figure 2.** (a) Schematic representation of the 3D printing process with the food ink; (b) Model of
 241 the 40-layers 3D scaffold, (c) As-printed 40-layers GB/XG scaffold and (d) GB/XG + 37.5 mM
 242 Ca^{2+} scaffold using 200 μm nozzle; (e) Representative optical top-view image of a GB/XG scaffold
 243 and (f) GB/XG + 37.5 mM Ca^{2+} scaffold. All the photos and images were taken at room
 244 temperature right after printing. Scale bar is 1 mm. The printing time for a 12 mm \times 12 mm scaffold
 245 is around 10 min.

246 **3.2. Rheological properties of food inks**

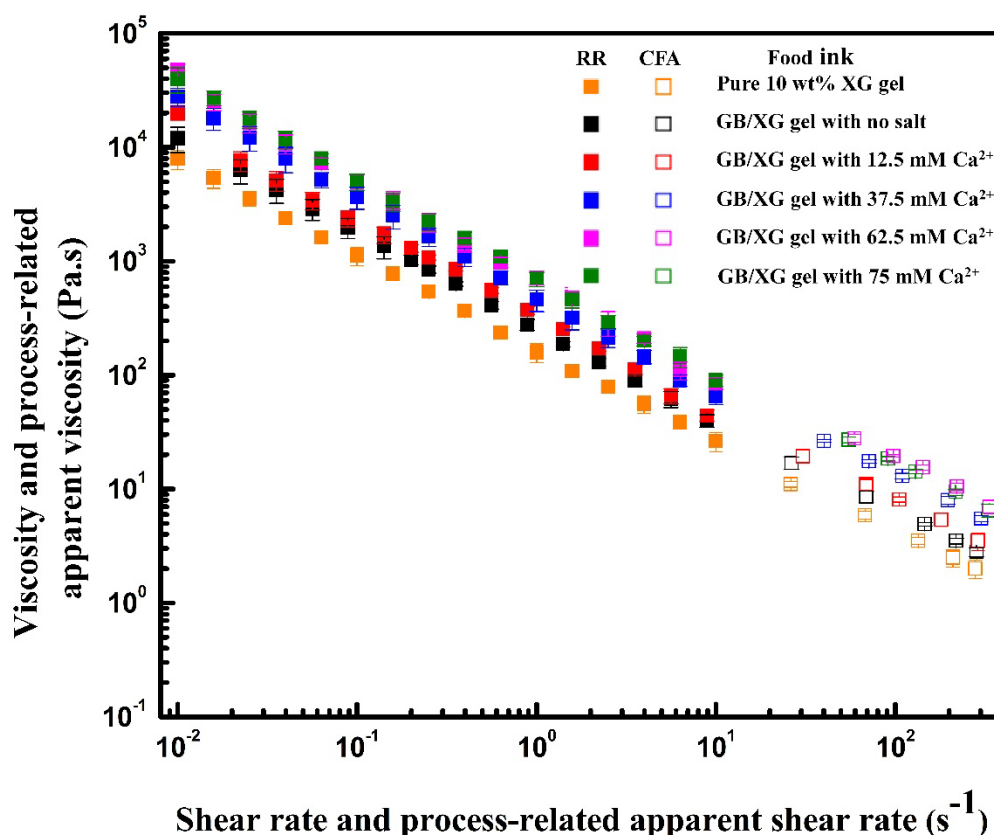
247 The rheological properties of the GB/XG ink are of critical importance since they are directly
 248 related to the extrusion process and ensure successful flow through the micronozzle and good post-
 249 deposition properties.

250 **Figure 3** shows the shear viscosity of neat XG and the 3 wt GB/10 wt% XG aqueous mixture with
251 different Ca^{2+} concentrations. The solid symbols in the low-to-moderate shear rate region (~ 0.01 -
252 10 s^{-1}) represent the steady-shear viscosity of the food inks measured by rheometer at room
253 temperature. The open symbols in the higher shear rate region ($> 10 \text{ s}^{-1}$) show the process-related
254 apparent viscosity of the food inks as a function of the process-related shear rate. These results
255 were calculated from capillary flow analysis (CFA) (Guo, et al., 2014) data obtained using a nozzle
256 diameter of $200 \mu\text{m}$ when extruding the material at room temperature. All food inks demonstrated
257 a shear-thinning behavior, which is favorable for the printing process. The viscosity decreases
258 significantly with increasing shear rate, with a relatively low power-law index (n) (**Table 2**),
259 typical of soft solids such as XG (J.A. Carmona, 2017). The data measured by rotational rheometer
260 (RR) is generally coherent with the data calculated from CFA. However, as seen in **Table 2**, n
261 values calculated from CFA are relatively higher than the results measured using the rheometer.
262 The decreased shear rate dependency indicates that the structure of the food inks is already rather
263 oriented or stretched in capillary flow (Ansari, Rashid, Waghmare, & Nobes, 2020; Drabek,
264 Zatloukal, & Martyn, 2018).

265 Compared with XG alone, the GB/XG food ink shows higher viscosity, which might be due to
266 their synergistic interactions, which benefits the gelation of hydrocolloids. The electrostatic
267 repulsions between GB and XG enhance the local concentration of XG. In addition, XG also
268 interacts with the negative patches on GB by electrostatic attractions (C.-S. Wang, et al., 2016).
269 As shown in **Table S1**, the viscosity increased by approximately 46 % for food ink with GB
270 addition at 1 s^{-1} . On the other hand, an increase in the viscosity of the GB/XG food ink was
271 observed in the presence of Ca^{2+} , which is probably due to the bridging effect of Ca^{2+} with XG
272 molecules (Freitas, Cortez-Vega, Pizato, Prentice-Hernández, & Borges, 2013; Mohammed,
273 Haque, Richardson, & Morris, 2007; Patel, Maji, Moorthy, & Maiti, 2020) and/or negative patches
274 on GB. With the increase of Ca^{2+} concentration, the viscosity at 1 s^{-1} of different food inks ranged
275 from 158 ± 30 to $720 \pm 91 \text{ Pa}\cdot\text{s}$ (**Table S1**). A maximum viscosity is obtained when Ca^{2+}
276 concentration is 62.5 mM . Further increasing the Ca^{2+} concentration to 75 mM does not lead a
277 significant change in the viscosity. Therefore, an appropriate improvement in viscosity benefits
278 the extruding ability of food ink and thus the printability.

279 **Table 2.** Power-law index of different food inks within two specific shear rate regimes (two
 280 methods)

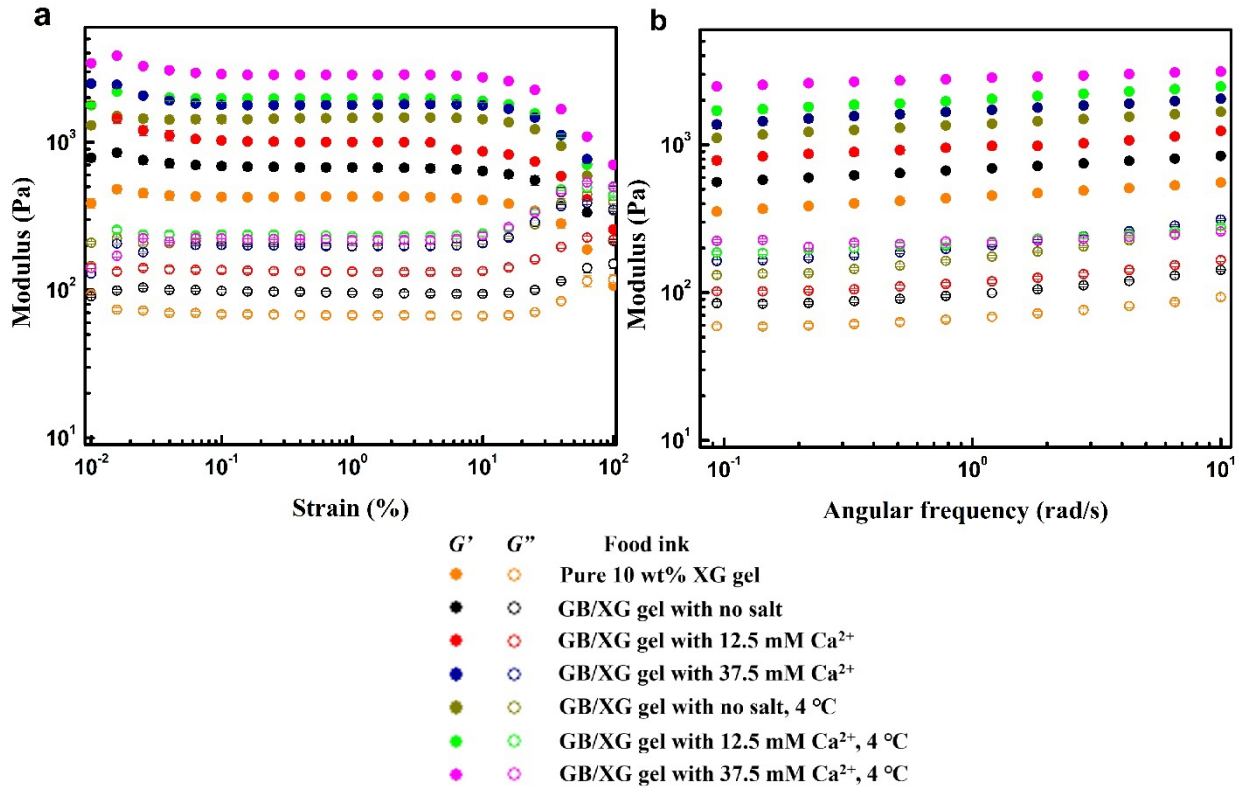
Food ink type	Power-law index (n)	
	Low to moderate shear rate (RR)	High shear rate (CFA)
Pure 10 wt% XG gel	0.18±0.02	0.27±0.07
GB/XG gel with no salt	0.16±0.03	0.25±0.07
GB/XG gel with 12.5 mM Ca ²⁺	0.14±0.03	0.24±0.04
GB/XG gel with 37.5 mM Ca ²⁺	0.12±0.03	0.22±0.02
GB/XG gel with 62.5 mM Ca ²⁺	0.12±0.02	0.21±0.03
GB/XG gel with 75 mM Ca ²⁺	0.12±0.02	0.21±0.03



281 **Figure 3.** Viscosity as a function of shear rate for XG and GB/XG food ink with different Ca²⁺
 282 concentrations at room temperature (22 °C) (solid symbols: data measured using a parallel plate
 283 flow geometry in steady shear (RR); open symbols: data obtained by capillary flow analysis (CFA)).
 284
 285 Linear viscoelastic properties of neat XG solution and GB/XG food ink with 0-37.5 mM Ca²⁺
 286 addition were also measured to infer the post-deposition quality, and the results are shown in
 287 **Figure 4.** For all food inks, the linear viscoelastic regime is identified in-between a strain

288 amplitude of 0.2-5% (**Figure 4a**). The storage modulus G' dominates the loss modulus in all the
289 investigated strain and frequency ranges (**Figure 4a and 4b**). In addition, G' of GB/XG food ink
290 without salt is almost 1.5 times larger than G' of neat 10 wt% XG. Adding Ca^{2+} to the ink leads to
291 an increase of G' , possibly due to the bridging effects of Ca^{2+} (Freitas, et al., 2013; Mohammed,
292 et al., 2007; Patel, et al., 2020), which enhances the gel strength of the gel formulation and also
293 allows the corresponding ink to retain a high shape fidelity after deposition (**Figure 2f**). Comparing
294 Ca^{2+} concentration ranging from 0 to 37.5 mM, G' gradually increased from 452 ± 26 to $1720 \pm$
295 68 Pa (380%) (**Table S2**). Similar observations of gel properties enhancement have been found
296 for the gelatin-transglutaminase (TG) system (Du, et al., 2021). The increases were almost 2 times
297 in G' at 1 rad/s and viscosity at 1 s^{-1} at low very content of transglutaminase (2 mg/mL) due to
298 strong chemical crosslinking between gelatin and TG as opposed to physical crosslinking between
299 gelatin and calcium ions. Regarding the power-law parameter a on frequency dependency of G'
300 (**Table S3**), it decreased when lowering the temperature, which resulted in a strengthening in gel
301 structure. The decrease in frequency dependence indicated a more solid-like behavior when
302 decreasing the temperature. Lowering the temperature to $4 \text{ }^\circ\text{C}$ favors the intra- and intermolecular
303 hydrogen bonding formation (Zhang, Wang, Wang, Han, & You, 2020), which boosts the
304 formation of stiffer structures, such triple helical structure of GB (Bello, Kim, Kim, Park, & Lee,
305 2020) and double helical structure of XG (Lei Wang, Xiang, Li, Zhang, & Bai, 2021). This results
306 in a higher G' and enhances the gel strength of the food ink. For example, the G' of the GB/XG
307 food ink with 12.5 mM Ca^{2+} addition at frequency of 1 rad/s is 2040 ± 97 Pa at $4 \text{ }^\circ\text{C}$, which is
308 about twice higher than the G' measured at room temperature (**Table S2**).

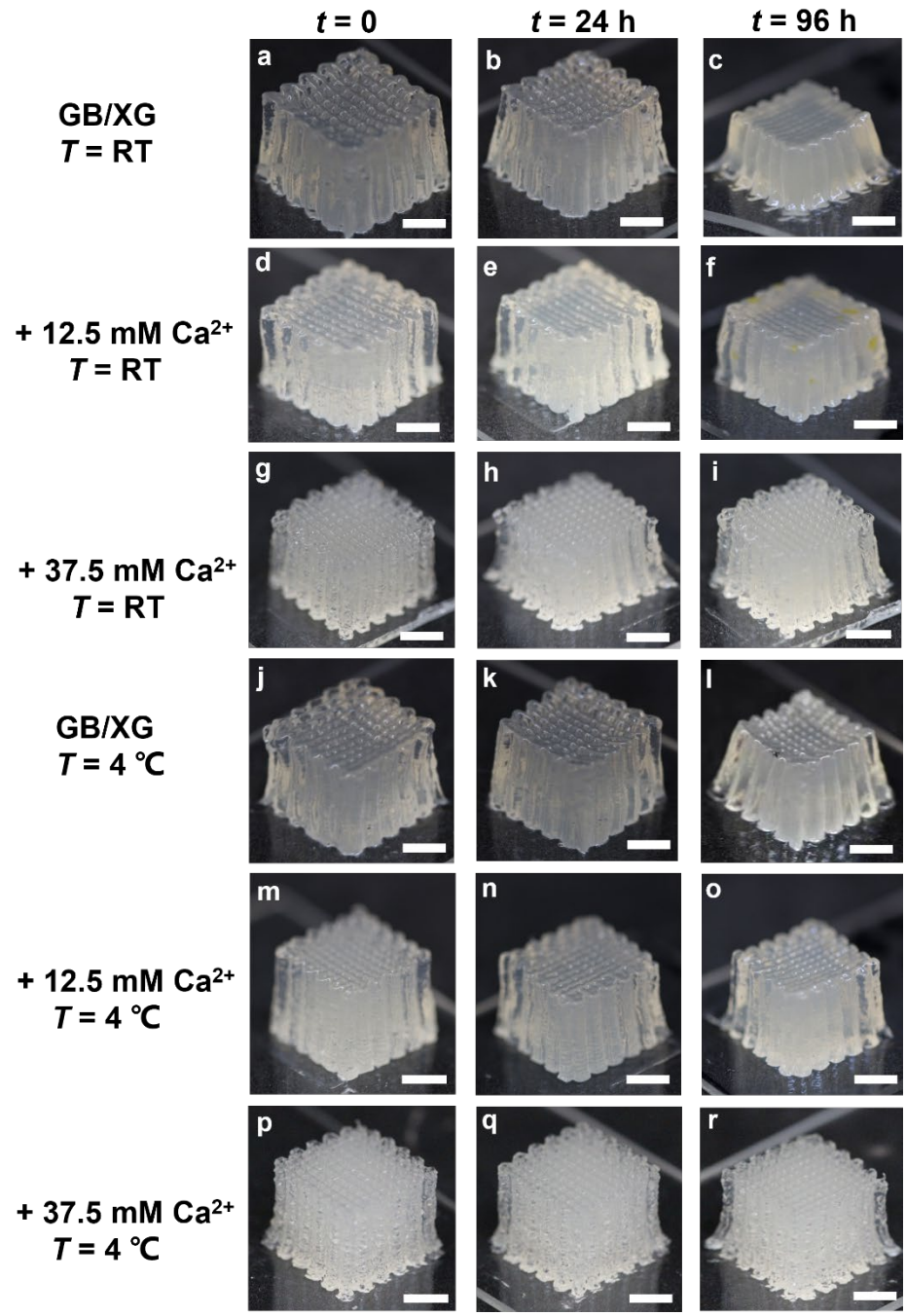
309 **Table S2** also summarizes the $\tan \delta$ of different food inks at an angular frequency of 1 rad/s. The
310 presence of GB in XG, the addition of Ca^{2+} and lower temperature resulted in lower $\tan \delta$ values,
311 which were in a range from 0.08 to 0.15. Those gels can be qualified as weak gels (Rao, 2014),
312 which benefited for the printing process.



313

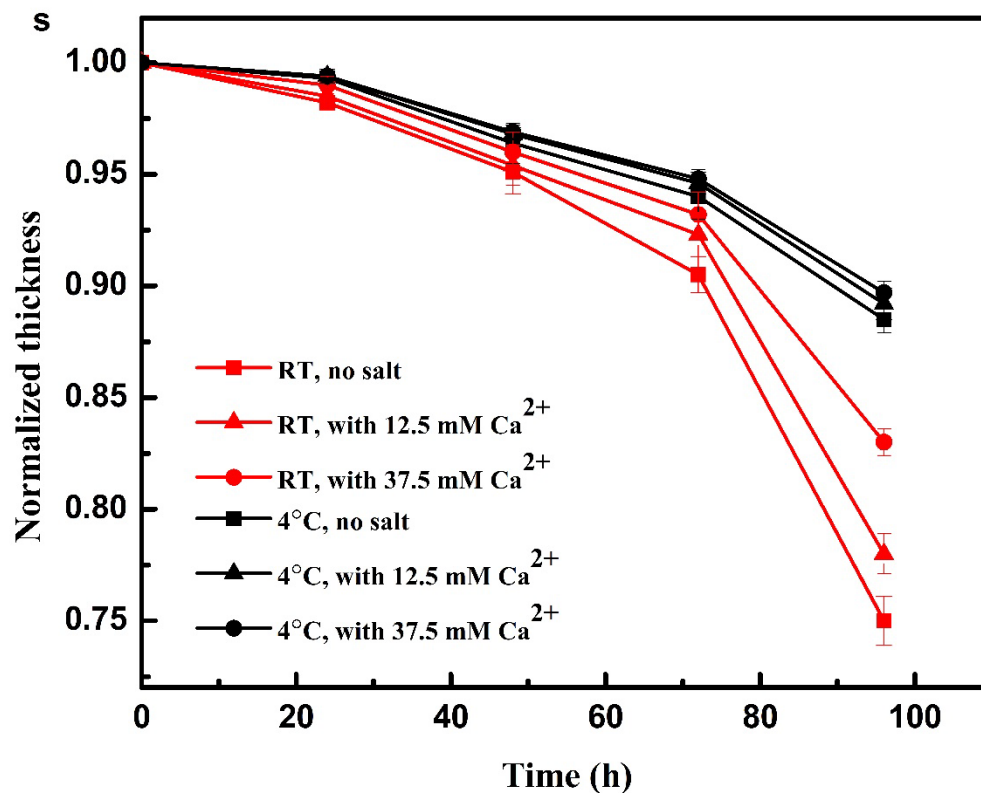
314 **Figure 4.** Storage modulus G' (solid symbols) and loss modulus G'' (open symbols) for XG gel
 315 alone and GB/XG mixed gels with different Ca^{2+} addition at room temperature (RT) and 4 °C: (a)
 316 as functions of strain amplitude (%) at $\omega = 1$ rad/s (b) as functions of angular frequency at
 317 strain = 1%.

318 **3.3 Dimensional stability of 3D-printed structures**



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321
 322 **Figure 5.** As-printed 40-layer scaffold using GB/XG mixed gels (a-c) with no salt addition (d-f)
 323 with 12.5 mM Ca²⁺ and (g-i) with 37.5 mM Ca²⁺ addition after printing 0, 24 and 96 h stored at
 324 room temperature (22 ± 1°C). As-printed 40-layer scaffold using GB/XG mixed gels (j-l) with no
 325 salt addition (m-o) with 12.5 mM Ca²⁺ addition and with 37.5 mM Ca²⁺ addition (p-r) after printing
 326 0, 24 and 96 h at stored at 4 °C. (s) Normalized thickness of the scaffolds fabricated using GB/XG
 327 food ink with 0, 12.5 mM and 37.5 mM Ca²⁺ addition stored at room temperature and 4 °C over a
 328 period of 96 h. Scale bars are 5 mm.

329 The dimensional stability of 3D-printed structures was investigated as it is essential for food
 330 storage. A 4 day duration was chosen as a goal for ready-to eat food storage. **Figures 5a-r** presents
 331 the structure images showing the effect of Ca²⁺ addition and storage temperature after printing on
 332 the dimension of as-printed 3D scaffolds with respect to time. In general, adding Ca²⁺ or decreasing
 333 temperature improves the dimensional stability. Printed scaffold with sharper outline can be
 334 obtained for the GB/XG ink with added Ca²⁺ (**Figure 5d and 5g**) compared to the salt-free ink
 335 (**Figure 5a**). Besides, shape retention improves when increasing Ca²⁺ concentration (**Figure 5d**
 336 **and 5g**). The same phenomena has been reported based on the gelation quality of calcium-

337 enhanced alginate gels (Dalheim, et al., 2019) and pectin gels (Vancauwenberghe, et al., 2017).
338 As the shape fidelity after deposition is closely related to the yield stress, the apparent yield stress
339 of different food inks was calculated based on the method by Bruneaux et al. (Bruneaux, et al.,
340 2008), and the results are shown in **Figure S2**. The apparent yield stress of food inks increases
341 with a linear concentration dependence on Ca^{2+} in the range of 0 to 62.5 mM, which explains the
342 better shape retention after printing. The high apparent yield stress is probably due to the reinforced
343 gel network with Ca^{2+} addition. The latter modifies the flow properties but also helps to induce
344 gelation (Freitas, et al., 2013; Mohammed, et al., 2007; Patel, et al., 2020) and maintain the 3D
345 structure during and after printing . On the other hand, lowering the storage temperature results in
346 good shape retention ability of the 3D printed scaffolds (**Figure 5m-5r**). The increased G' at low
347 temperature from the rheological results (**Figure 4**) can also justify the superior shape retention.
348 Besides, low temperature slows down the solvent evaporation, hence a higher shape fidelity of the
349 hydrogel is retained.

350 **Figure 5s** shows the dimensional stability of the as-printed scaffolds by monitoring the evolution
351 of the relative thickness variation to quantify the structure slumping. All the samples retain at least
352 75 % of their initial thickness after 96 h, indicating a satisfactory shape stability with food ink
353 using the GB/XG pair. Comparably, thickness reduction using the food ink with 12.5 mM and 37.5
354 mM Ca^{2+} is 3% and 8% slower than the one with salt-free ink. In addition, the thickness reduction
355 for scaffolds with both GB/XG and GB/XG/ Ca^{2+} food ink is quite similar at 4 °C, showing around
356 11% reduction after 96 h, indicating that the storage temperature effects dominate dimensional
357 stability compared to the salt effect.

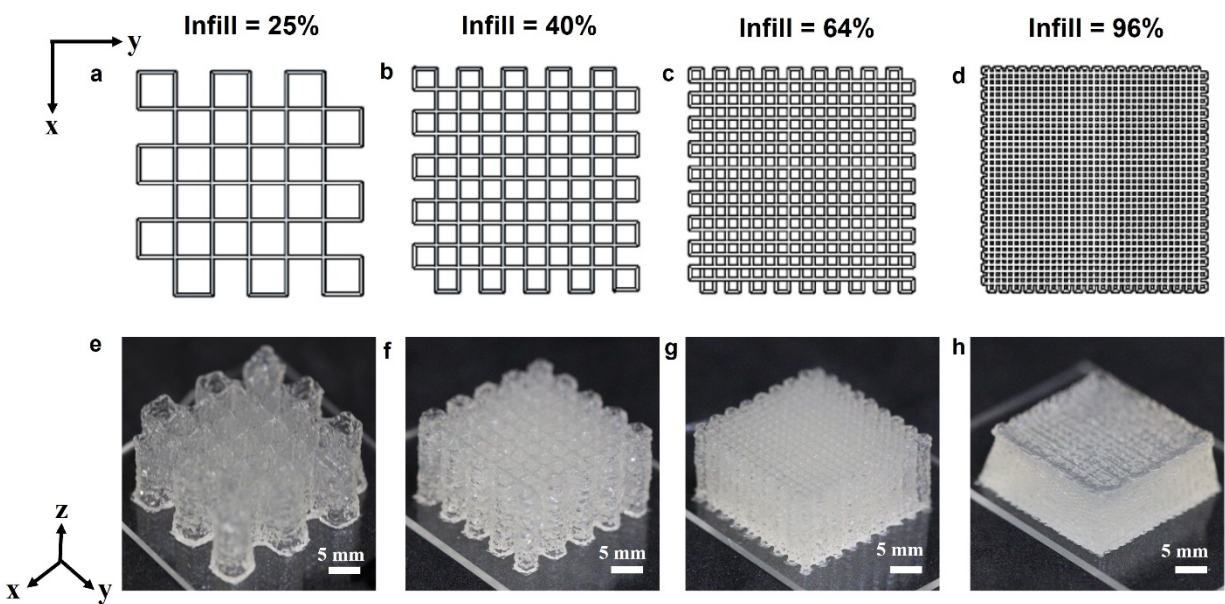
358 Overall, the dimensional stability agrees well with the fact that the rheological properties are
359 enhanced when adding Ca^{2+} or decreasing temperature.

360 ***3.4 Compression testing for textural profile analysis (TPA)***

361 TPA is one of the most common measurement method for describing the characteristics of gels
362 (Tao, et al., 2021), which includes hardness, adhesiveness, cohesiveness, springiness and
363 gumminess. These characteristics are strongly related with the sensory perception of food (Çakır,
364 et al., 2012). Scaffolds with different infill percentage were first printed (**Figure 6**) and were then
365 tested for TPA. The calculated textural parameters are shown in **Figure 7 (Table S4** for more

366 details) in order to clarify the effect of infill percentage of 3D scaffolds, Ca^{2+} concentration in the
367 GB/XG food ink and storage temperature of printed scaffolds on basic textural properties.

368 Hardness is related to the sensory perception of the first bite in the mouth, which is the most direct
369 reaction mouthfeel (Chandra & Shamasundar, 2014). It is represented by the peak force of the first
370 compression cycle. The effects of infill percentages, salt concentrations and storage temperature
371 on the hardness of the samples are shown in **Figure 7a**. Increasing the infill percentage causes an
372 increase in the hardness. Specifically, the samples with 96% infill percentage have a significant
373 higher hardness value compared to the ones with lower infill percentage ($p < 0.05$). The scaffolds
374 with 25% infill percentage collapsed easily upon compressing because of the small contact
375 filament area with the probe and thus high pressure. One interesting point to mention here is that
376 the infill percentage can be well controlled during the 3D food model design process, so it can
377 meet any specific requirements in terms of food texture, highlighting the potential of 3D printing.
378 A significant difference in hardness was also observed for the samples containing different Ca^{2+}
379 concentration ($p < 0.05$). GB/XG food ink with 62.5 mM Ca^{2+} addition has the highest hardness
380 value of 57.3 g. Regarding the effect of storage temperature, it has an inverse relationship with
381 hardness. That is, a decrease in storage temperature leads to an increase in the hardness, which is
382 probably due to the formed stiffer structure, as discussed above.



383

384 **Figure 6.** (a-d) Top-view models of scaffold with different infill percentage. (e-h) Representative
385 images of 3D printed scaffold with different infill percentage using GB/XG with 37.5 mM Ca²⁺
386 food ink at room temperature right after printing. The printing times for scaffolds with different
387 infill percentages were varying between 8 and 30 min.

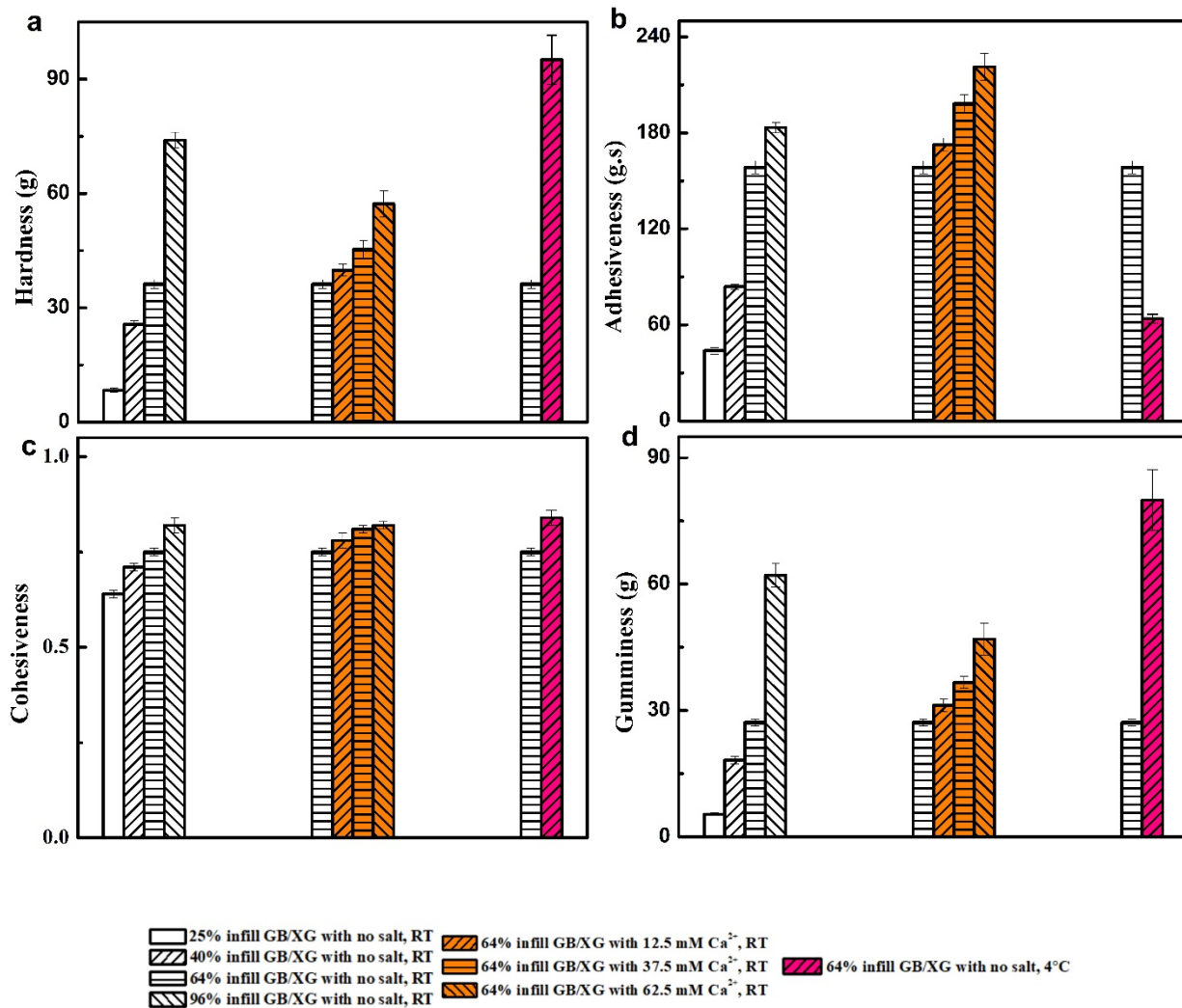
388 When retracting the probe, a negative force arises from the attraction between the sample surface
389 and the probe. Adhesiveness is defined as the negative area of force when the probe withdraws
390 after the first compression. It can be interpreted as a sticky mouthfeel (Siamand, Deeth, & Al-
391 Saadi, 2014). Appropriate adhesiveness benefits the taste and flavor releasing. However, too strong
392 adhesiveness may cause the food sticking to the inner packaging, thus a structural damage when
393 removing the packaging (Chandra, et al., 2014). The adhesiveness for samples with different infill
394 percentages, salt concentrations and storage temperatures are shown in **Figure 7b**. The scaffold
395 with the densest structure has a significant higher adhesiveness of 183 ± 3 g.s, because of the larger
396 contact surface ($p < 0.05$). Adding Ca²⁺ to the ink also results in an increase of adhesiveness, with
397 the sample containing 62.5 mM having a significant higher value ($p < 0.05$), which might be
398 explained by the increased G'' after Ca²⁺ addition. Similar observations for XG-based food gels
399 have been reported before (Yang, Tsai, Jiang, & Hua, 2021). As for storage temperature, lower
400 adhesiveness is observed at 4 °C because the structures containing GB become more elastic, as
401 mentioned before.

402 The cohesiveness indicates the strength of internal bonds making up the body of food and the
403 degree to which a food can be deformed before it breaks. It is defined as the ratio of the positive
404 force area during the second compression to that of the first compression (Siamand, et al., 2014).
405 Cohesiveness comparison of infill percentages, salt concentrations and storage temperatures
406 effects are shown in **Figure 7c**. Samples with higher infill percentage shows stronger cohesiveness
407 because of a relatively more stable structure against compression. The higher apparent yield stress
408 for the food ink with higher Ca²⁺ provides better resistance against the deformation and thus
409 benefits the cohesiveness of the printed scaffold. Moreover, food ink at low temperature
410 demonstrates a higher gel strength, which translates into a larger tensile strength and higher
411 cohesiveness of the printed scaffolds.

412 Gumminess is defined as the product of hardness and cohesiveness, which indicates the required
413 energy to disintegrate food for swallowing (Nishinari & Fang, 2018). It is another important

414 textural characteristics, especially for semisolid food with a low degree of hardness and high
415 degree of cohesiveness (Chandra, et al., 2014). In our case, gumminess exhibits a similar trend to
416 hardness, regardless of the effects of infill percentage, salt concentration and storage temperature,
417 as shown in **Figure 7d**. This indicates that the gumminess of printed samples is dominated by their
418 hardness (Yang, et al., 2021).

419 Overall, food texture can be tuned by the rheological properties of the food ink, infill percentage
420 during 3D printing and storage temperature. By carefully controlling the food ink properties and
421 the designed form, it is possible to create group-specific food constructs according to their
422 requirements. Food model designed with 100% infill are more popular in visual sight, while it is
423 more acceptable in terms of mouthfeel with 25% infill (Mantihal, Prakash, & Bhandari, 2019).
424 Some special groups of people such as dysphagic patients with swallowing difficulties are usually
425 not able to consume food with high adhesiveness (Bychkov, Reshetnikova, Bychkova,
426 Podgorbunskikh, & Koptev, 2021). 3D printing shows its ability control food properties for a large
427 range of needs. For food applications, the 3D printed scaffolds using our food inks have a similar
428 range of adhesiveness and cohesiveness as cheddar (Y. Zheng, Liu, & Mo, 2016), while their
429 hardness and gumminess values can be referred to that of Jello™ (T. K. Kim, et al., 2020) and
430 pudding (Park, Lee, Yoo, & Nam, 2020; J. Zheng, Wu, Dai, Kan, & Zhang, 2017). This shows the
431 great potential to use the mixture of GB and XG to simulate the texture of soft food, well in
432 agreement with a previously published work by Cohen et al.(Cohen, et al., 2009).



433

434

435 **Figure 7.** Ca²⁺ concentration, infill percentage and storage temperature effects on (a) hardness (b)
 436 adhesiveness (c) cohesiveness and (d) gumminess.

437

438 4. Conclusions

439 We successfully 3D printed food-grade scaffolds with ink composed of GB mixed with XG. The
 440 GB/XG ink exhibits a shear-thinning behavior, which allows the food ink to be easily extruded
 441 from the printer nozzle. GB is a key component as it endows the ink with improved rheological
 442 properties and the scaffold with better dimensional stability compared to that of XG alone. The
 443 addition of Ca²⁺ to GB/XG ink improves the printing results due to its bridging effect. Scaffolds
 444 exhibited the best structure and shape retention using food ink with the addition of 37.5 mM Ca²⁺.

445 However, ink with 62.5 mM Ca^{2+} can cause irregular filaments and structure defects possibly
446 because it causes over-gelation of the food ink. All printed scaffolds using GB/XG can retain more
447 than 75% of their original thickness after 96 h of storage at room temperature, regardless of the
448 presence of Ca^{2+} , exhibiting a high shape fidelity retention. Scaffolds with both GB/XG and
449 GB/XG/ Ca^{2+} food ink show the lowest thickness reduction - around 11% - after 96 h at 4 °C.
450 Finally, the textural properties of the printed food scaffold can be modulated by adding Ca^{2+} to the
451 ink, controlling the infill percentage through the 3D printing process and using different storage
452 temperature after printing. This provides the possibility to use the 3D printing technique to shape
453 group-specific food formulations according to different needs, such as those of elderly people or
454 dysphagic patients.

455

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